

available at www.sciencedirect.com







Commentary

The powerful cardioprotective effects of urocortin and the corticotropin releasing hormone (CRH) family

Sean M. Davidson a,*, Aneta E. Rybka b, Paul A. Townsend b

^a The Hatter Cardiovascular Institute, University College London Hospital and Medical School, 67 Chenies Mews, London WC1E 6HX, UK ^b Human Genetics Division, Duthie Building MP808, School of Medicine, University of Southampton, Southampton General Hospital, Southampton SO16 6YD, UK

ARTICLE INFO

Keywords: Urocortin Cardiovascular Cardioprotection Heart failure Ischemia and reperfusion

ABSTRACT

The urocortins are members of the corticotropin releasing hormone (CRH) family of peptide hormones. The archetypal member of this family, CRH, plays an important role in regulating thermogenesis and homeostasis by acting centrally and systemically in target organs via its two receptors CRH-R1 and CRH-R2. However, by virtue of their much greater relative affinity for CRH-R2, the physiological effects of the urocortin peptides are largely restricted to peripheral organs such as the heart. A powerful cytoprotective effect of urocortin peptide administration against ischemia and reperfusion injury has been demonstrated in isolated cardiomyocyte models, as well as in the intact heart both in vitro and in vivo. Extremely promising data has shown the beneficial effect of treating pacing-induced heart failure in sheep with urocortin molecules. Though the efficacy and specificity of these molecules in humans is not yet established, molecular dissection of the cytoprotective pathways activated by urocortin peptides suggests that the beneficial effects may be separable from potentially deleterious effects.

© 2008 Elsevier Inc. All rights reserved.

1. Introduction

Urocortin (Ucn 1) is a 40-amino acid member of the corticotropin releasing hormone (CRH) family of peptide hormones [1] (see Fig. 1 for the urocortin peptide structures). The archetypical member, CRH itself, is produced in the brain in response to stress, and can affect behaviour by acting locally on the brain, as well as by affecting autonomic responses in peripheral organs. The various peptides of the CRH family appear to have overlapping roles in various tissues including the placenta, immune system, digestive system, central nervous system and cardiovascular system, with their relative importance in each system depending on their site of production, plasma distribution and specific receptor affinity.

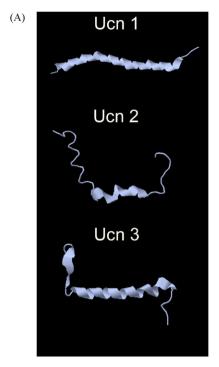
This review is intended as a synthesis of current knowledge on the role of CRH-family proteins in the heart, particularly their therapeutic potential in terms of cardioprotection. We will also cover their important centrally mediated physiological effects, which must be taken into account if Ucn 1 or one of its homologs is to ever be developed as pharmaceutical agent. The reader is referred elsewhere for reviews on the role of the CRH family in other tissues [2].

2. The CRH-peptide family and their receptors

There are two mammalian genes encoding CRH receptors, both of which are expressed in the brain, and both of which

^{*} Corresponding author. Tel.: +44 207 380 9781; fax: +44 207 380 9505. E-mail address: S.Davidson@ucl.ac.uk (S.M. Davidson).

0006-2952/\$ – see front matter © 2008 Elsevier Inc. All rights reserved. doi:10.1016/j.bcp.2008.08.033



(B)	CRH	SEEPPISLDLTFHLLREVLEMARAEQLAQQAHSNRKLMEII
(2)	Ucn1	-DNPSLSIDLTFHLLRTLLELARTQSQRERAEQNRIIFDSV
	Ucn2	IVLSLDVPIGLLQILLEQARARAAREQATTNARILARV
	Ucn3	TKFLSLDVPTNIMNLLFNIAKAKNLRAOAANAHLMAOI

)		HGNC	mRNA	PID	PDB
	CRH	2355	NM_000756	P06850	-
	Ucn 1	12516	NM_003353	P55089	2RMF
	Ucn 2	18414	NM_033199	Q96RP3	2RMG
	Ucn 3	17781	NM 053049	Q969E3	2RMH

Fig. 1 – (A) Peptide structures obtained from PDB file co-ordinates [12]. (B) Sequence alignments of human CRH-family peptides created using ClustalW2, where amino acids coloured red are nonpolar or hydrophobic; blue are acidic; magenta are basic; green are polar or non-charged. (C) Database references for human Ucn 1, Ucn 2, and Ucn 3. HGNC: the unique reference identity given to the human gene by the HUGO gene committee; mRNA: the Genbank reference id; PID: the EBI UniProt identifier; PDB: the 3D structural data identifier. (For interpretation of the references to color in the figure caption, the reader is referred to the web version of the article.)

can bind all members of the CRH family albeit with different affinities. CRH-R1 is not detected in the heart [3], but CRH-R2 is highly expressed in cardiomyocytes, as well as in the gastrointestinal tract and epididymis [3–6]. There are two splicing variants of CRH-R2 in the heart—while CRH-R2 α has been detected in all chambers of the human heart, CRH-R2 β seems to be restricted to the left atrium [3]. CRH-R2 β is also found in endothelial and smooth muscle cells of the systemic vasculature [5].

(C)

Expression of Ucn 1 has been detected in all four chambers of the human heart, although CRH is absent [3]. Since Ucn 1 is about 10-fold more potent than CRH at binding and activating CRH-R2 [1], Ucn 1 was anticipated to have effects on the heart—as was later borne out experimentally (see below). However, since it also binds with strong affinity to CRH-R1 it has concurrent effects in the brain.

Urocortin 2 (Ucn 2, also called stresscopin-related protein or SRP [7], a 38-aa peptide originally detected in mouse brain, binds CRH-R2 selectively and with high affinity [8]. Ucn 2 can

be detected by RT-PCR in various tissues including heart and muscle [7]. Apparently because of its low affinity for CRH-R1, administration of the peptide to mice does not increase gross motor activity, but it does still have effects on central autonomic and appetite control [8].

Urocortin 3 (Ucn 3, also called stresscopin or SCP [7]) was identified on the basis of its sequence homology to other CRH-family members, and encodes a 38-aa mature peptide that is 90% homologous in mouse and humans. It is widely expressed, particularly in areas of the brain, the digestive system, and cardiac and skeletal muscle [7,9]. Ucn 3 is also strongly expressed in pancreatic β -cells, where it appears to act locally to augment insulin production—consequently mice lacking Ucn 3 are relatively protected from the harmful metabolic consequences of a high-fat diet, maintaining better glucose tolerance [10]. Additionally, it can be detected in normal human plasma [11]. Like Ucn 2, Ucn 3 is selective for CRH-R2 [9].

The NMR structures of human Ucn 1, Ucn 2 and Ucn 3 in solution have been determined—they are primarily alpha-

helical, with a small kink or turn at residues 25–27, resulting in a helix-turn-helix motif believed to be important for their binding to and activation of CRH receptors [12]. An important tool in examining the role of CRH-family peptides is the artificially synthesized antagonist, alpha-helical CRH-(9–41), which potently inhibits the effects of CRH-family peptides [13].

3. The role of the CRH-peptide family in homeostasis

CRH has an interesting and very important role in regulating thermogenesis and energy homeostasis. Mice deficient for CRH-R2 have significantly elevated basal levels of thermogenesis in brown adipose tissue, and exhibit behavioural changes indicating a loss of body heat [14]. CRH can act directly on skeletal muscle to stimulate thermogenesis, possibly via substrate cycling between de novo lipogenesis and lipid oxidation. This effect can be blocked by inhibitors of phosphatidylinositol 3kinase or AMP-activated protein kinase [15]. Ucn 2 is proposed to function as a local negative regulator of glucose uptake in skeletal muscle, since mice deficient in Ucn 2 or administered with a CRH-R2 antagonist exhibit increased insulin sensitivity and are protected against fat-induced insulin resistance [16]. Other homeostatic effects of Ucn may be centrally mediated, for example intracerebrovascular injection of an alpha-helical CRH-(9-41) into rats blocks the pyrogenic and thermogenic effects of interleukin 1β [17].

The effects of CRH-family peptides on anxiety and activity seem to be mediated by CRH-R1 whereas effects on appetite are mediated via CRH-R2 [18], therefore the different effects of Ucn [19] and CRH may be explained by different receptor binding profiles [20]. Other centrally mediated effects of Ucn include a role in fluid homeostasis [21] and in osmoregulation, reflecting the role of orthologous proteins in insects [22] Table 1A.

4. The CRH-peptide family and disease

Due to the wide range of expression of the urocortins and their receptors, and their powerful effects on homeostasis, their potential involvement in various diseases has been investigated. It has been found that, unlike CRH, the urocortins do not increase corticosterone secretion and seem not to have a physiological role in regulating the hypothalamic-pituitary-adrenal (HPA) axis [18].

CRH-family peptides also exhibit an anti-inflammatory effect that appears to involve a direct effect on macrophages [23] but may also be stress-related. Consequently, the potential for CRH-1 receptors as a therapeutic target for inflammatory diseases such as irritable bowel syndrome has been investigated [24]. An increase in Ucn expression is detected in lung tissue of rats with allergic asthma [25], although whether it is playing a pro- or anti-inflammatory role in this case is not known.

5. Cardiovascular effects of the CRH-peptide family

CRH is unlikely to have major direct effects on the heart, since plasma concentrations are very low and it is not highly expressed locally. In contrast, as mentioned above, Ucn 1, Ucn 2 and Ucn 3 are expressed in the heart. Expression and secretion of Ucn 1 increases in isolated rat cardiomyocytes exposed to simulated ischemia [26]. Furthermore, cardiac release of Ucn 1 occurs in the absence of tissue necrosis when rats are exposed to sublethal ischemia and reperfusion injury, reaching ${\sim}8~{\rm pM}$ in the plasma, compared to undetectable levels in control rats [27]. Arterial plasma Ucn 1 levels measured $15.2\pm0.5~{\rm pM}$ in normal sheep and increased significantly following development of heart failure to

Table 1A – General physiological effects of urocortins						
Species	Work performed	Relative functional effects	References			
Rat	Food and water intake suppression Gastric motility inhibition Autonomic control of gastric emptying	Regulation of caloric intake	[19,85]			
Rat	Contribution to an increase in lung vascular permeability through activation of CRH receptor	May act as an autocrine and paracrine proinflammatory factor in lungs	[86]			
HF sheep (experimental heart failure)	Urine volume increase	Effects on renal hemodynamics	[35]			
Human (synovial tissue of patients with rheumatoid arthritis RA)	Possible inhibitor of extravesation	Possible suppression of the inflammation in RA	[87]			
Human (normal cycling human ovaries)	Most likely inhibition of ovarian steroidogenesis in the process of luteal degeneration	May act as autocrine and/or paracrine regulator in the ovarian corpus luteum	[88]			
Mouse and rat (mesangial cells)	Suppresses production of ROS in endothelial cells and sustains endothelium-dependent relaxations of coronary artery	Could significantly ameliorate diabetic nephropathy	[89]			
Mouse	Growth inhibition of hepatocellular carcinoma (HCC) and reduction of tumor microvessel density	Reported to be a tonic suppressor of vascularization, implying its role in tumor angiogenesis	[90]			

Species	Model	Effects observed	References
Mouse	in vitro in vivo	→ Protection of cardiac myocytes from cell death induced by hypoxia ↓ inotropic, lusitropic and systemic arterial load on the LV myocardium ↑ cardiac output	[47] [38]
	in vivo (dilated cardiomyopathy) in vivo (conscious rats)	Significant improvement in HF ↓ blood pressure ↑ heart rate	[1]
Rat	in vivo (anesthetized rats)	↓ arterial pressure ↓ cardiac output ↑ heart rate ↑↑ cardiac contractility	[85]
	in vitro	Protection of cardiac myocytes from cell death induced by hypoxia	[45,46,49,55–57, 59,60,66]
	Langendorff	Protection against ischemia and reperfusion injury	
Sheep	in υiυο	↑ arterial pressure ↑ cardiac output ↑ heart rate ↑ coronary blood flow ↑↑ cardiac contractility	[32]
	in vivo (experimental heart failure)	↓ arterial pressure ↑ cardiac output ↓ blood pressure	[35–37]
	in vivo (heart failure)	↑ cardiac output ↑ left ventricular ejection fraction ↓ arterial pressure	[43]
Human	in vivo (healthy individuals)	↑ cardiac output ↑ heart rate ↑ left ventricular ejection fraction	[44]

 19.1 ± 1.6 pM [28]. Similarly, Ucn 1 levels are increased from \sim 19 pM in normal human males to \sim 50 pM in those with systolic heart failure [29]. An immunohistochemical study also demonstrated increased immunoreactivity for Ucn 1 in the left ventricular myocytes of 9 dilated cardiomyopathic hearts compared to 5 normal human hearts [5]. Circulating Ucn 3 in normal human plasma has been reported at a level of \sim 50 pM [11] (Table 1B).

Ucn 1 has direct effects on the heart, increasing inotropy and coronary vasodilation in the isolated rat heart after administration of a bolus dose estimated to have attained a plasma concentration of >200 nM [30], and these effects appear to be mediated by CRH-R2 since the response is absent in mice lacking this receptor [31]. Administration of a 100 µg Ucn 1 but not CRH potently elevates cardiac contractility in sheep for up to 24 h [32]. Furthermore, intravenous administration of lower doses of Ucn 1 (2.5-10 μg) was recently shown to induce potent inhibition of sympathetic nerve activity to the heart despite being below the threshold for direct actions of Ucn1 on the myocardium [33], suggesting Ucn 1 has a role in normal cardiac homeostasis, and may have further beneficial effects in heart failure via indirect mechanisms. Indeed, endogenous urocortins have been shown to have a protective role in pacing-induced heart failure in experiments in which a CRH-R2 antagonist was administered

The group of Richards in Christchurch, New Zealand, have made great inroads into exploring the putative benefits of Ucn and related peptides in treating heart failure. Their initial studies of intravenous administration of Ucn 1, Ucn 2 and Ucn 3 in sheep with pacing-induced heart failure demonstrated dramatic improvements in cardiac output, peripheral resistance, left atrial pressure, mean arterial pressure, renal function and suppression of detrimental neurohormonal circulation [35–37], though the relative involvement of direct and indirect effects of Ucn is not known. Thirty minutes after bolus administration of 10 µg, 50 µg, or 100 µg, they measured increases from the control plasma concentration of Ucn 1 of $12.3 \pm 1.3 \, pM$ up to $219 \pm 19 \, pM$, $1777 \pm 91 \, pM$ and 4142 ± 175 pM, respectively [35], with most significant cardiac effects occurring after 50 μg or 100 $\mu g.$ In mice treated with Ucn 2, these effects have been shown to be independent of β adrenergic receptors [38]. The vasodilatory but not other cardiac effects of Ucn 1 and Ucn 2 involve nitric oxide [39,40]. The more rapid onset of action of Ucn 3 and its shorter duration of action may be a reflection of different pharmacokinetic profiles [37].

Importantly, there does not seem to be a desensitization effect to prolonged Ucn administration, at least over 4 days of intravenous infusion of Ucn 1 (0.3 $\mu g/(kg\,h)$, attaining up to 8000 pM plasma Ucn 1) in sheep with pacing-induced heart failure, with sustained beneficial effects observed at the end of the experiment [41]. Interestingly, potential "side-effects" such as food intake suppression and stress hormone production appeared to be transient and diminished after 2 days [41]. Furthermore, Ucn 1 treatment commencing from the onset of cardiac overload was able to repress progression to overt heart failure, supporting its therapeutic use early in the disease [42].

Recently, the effect of Ucn 2 administration to human heart failure patients has been examined. After 1 h intravenous infusion of 25 μ g or 100 μ g Ucn 2 (attaining plasma concentrations of ~114 pM and ~440 pM vs. 19 pM in those receiving placebo), there was a significant increase in cardiac output and left ventricular ejection fraction and decreased mean arterial pressure, systemic vascular resistance, and cardiac work, with no effect on hormone levels observed [43]. This is similar to the effects observed by the same investigators in healthy humans, although in that case subtle renal effects and activation of plasma renin, angiotensin II, and norepinephrine (at high-dose only) were also observed [44].

The CRH family is highly cardioprotective, with an ED_{50} in the nanomolar range in various experimental systems. Treatment with any of the urocortin peptides in vitro protects isolated neonatal and adult rat cardiomyocytes from ischemia and reperfusion injury [45–47]. In addition, they have been shown to be protective in the isolated perfused rat heart [27,45,47,48], even when present only during reperfusion [49]. An extensive variety of peptides are known to activate common cardioprotective pathways after binding their respective G-protein-coupled receptors, and these same pathways are believed to be involved in other mechanisms of cardioprotection such as ischemic preconditioning and

postconditioning (reviewed in [50–52]). In order to understand the powerful cardioprotective mechanism of the CRH-family peptides, their signal transduction pathways have been extensively investigated.

6. Signal transduction mechanisms of the CRH-peptide family

The cardioprotective effects of urocortins against ischemia and reperfusion injury involve the activation of several signalling pathways that have both cytoplasmic and mitochondrial targets and culminate in the alteration of cellular metabolism and the modulation of apoptosis (see Fig. 2). Although most of this signalling pathway has been investigated using Ucn 1, the other members of the family are believed to act similarly by virtue of their interaction with the same cell surface receptors. By initially binding to G protein-coupled receptors, Ucn 1 initiates a complex signalling cascade involving activation of phosphoinositide-3-kinase (PI3K), protein kinase A (PKA), protein kinase B/Akt (Akt), protein kinase C (PKC) and mitogen-activated protein kinases (MAPKs) amongst other signalling pathways [53]. Activation of these kinase pathways alters the activity of various channels

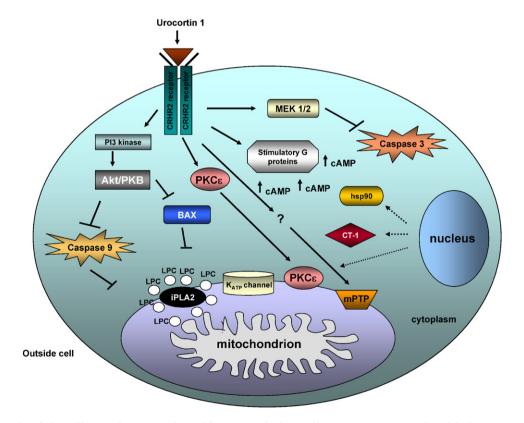


Fig. 2 – Schematic of signalling pathways activated by urocortin in cardiomyocytes. Urocortin 1 binds to CRH-R2 receptor, which causes the activation of stimulatory G proteins and results in activation of adenylyl cyclase and increased levels of cyclic adenosine monophosphate (cAMP). In parallel, mitogen-activated kinase p42/p44 (MAPK), phosphoinositide-3 kinase (PI3K) signalling pathways and protein kinase C epsilon (PKC ϵ) are activated. All three pathways have been reported to play a crucial role in cardioprotection. Downstream, late effects following urocortin exposure include increases in the expression of heat shock protein 90 (HSP90), cardiotrophin-1 (CT-1), K_{ATP} channel components, and PKC ϵ , in addition to a decrease in expression of the calcium insensitive phospholipase K_{C} (iPLA K_{C}). Opening of the mitochondrial permeability transition pore (MPTP) may be inhibited via PKC ϵ , or indirectly by decreasing levels of oxidative stress (see text).

including the mitochondrial permeability transition pore (MPTP), which is centrally involved in the induction of cell death [45,54]. In addition to acute effects on different kinase pathways urocortin peptides have more chronic, modulatory activity as shown by transcriptional and translational effects on mitochondrial ATP-sensitive potassium channel (K_{ATP}), calcium insensitive phospholipase A_2 (iPLA $_2$) and protein kinase C epsilon (PKC $_2$), all three intimately involved in cardioprotection [55–58].

Considerable data suggests that Ucn 1 acts via a p42/p44 MAPK (ERK1/2)-dependent signalling pathway in protecting both in vitro primary cell models and ex vivo and in vivo rodent heart from reperfusion injury. The extracellular signal-related kinases (ERK) belong to one sub-family of MAP kinases and consist of 42- and 44-kDa kinases. Their phosphorylation and activation is mediated by the MAP kinase MEK1/2 following Ucn 1 treatment [59,60]. Pharmacological "proof of concept" studies using the MEK1/2 inhibitors PD98059 and U0126 have demonstrated that Ucn 1-mediated cardioprotection requires activation of this signalling pathway. Specifically, in primary isolated cardiac myocytes, treatment with PD98059 abolished cytoprotection (in both ischemia and reperfusion) produced by Ucn 1 as assayed by trypan blue exclusion, annexin V staining and TUNEL labeling [48,59,60]. Moreover, inhibition of the MEK1/2 pathway by PD98059 decreased the ability of Ucn 1 to reduce infarct size during ischemia/reperfusion in an ex vivo perfused rat heart model as well as in vivo [59]. Importantly, similar findings were also reported for Ucn 2 and Ucn 3 [47]. Clearly, the p42/p44 MAPK pathway, which is activated by MEK1/2, seems to play a significant role in cardioprotective effect of Ucn.

Equally important, however, is phosphatidylinositol 3-OH kinase (PI3K) and one of its downstream effectors, protein kinase B (Akt), both of which are crucial for Ucn-stimulated increase in survival of cardiomyocytes [61]. Phosphorylation on both Thr-308 and Ser-473 as well as direct interaction with the PI3K lipid product (phosphatidylinositol-3 and 4-biphosphate) is required for Akt activation. Akt mediates cell survival in turn by phosphorylating the serine residues of a number of proteins including the pro-apoptotic protein BAD. This results in it binding to the 14-3-3 protein, thereby preventing it from interacting with mitochondria, and thus conferring resistance against apoptosis [62]. Both MEK1/2 and PI3K are responsible for preventing procaspases 9 and 3 from being cleaved into their active forms [63,64]. Furthermore, chemical inhibitors of the PI3K pathway, such as wortmannin and LY294002, have been reported to block Ucn 1-mediated cardioprotection in both neonatal and adult cardiomyocytes [61]. In this study, forced overexpression of active PI3K prevented the death of cardiomyocytes in response to simulated hypoxia and reoxygenation. Both Ucn 2 and Ucn 3 have also been demonstrated to act through the PI3K pathway [47].

In addition to its protective effects, urocortin treatment of both neonatal and adult rat cardiomyocytes has been shown to stimulate the secretion of atrial natriuretic peptide (ANP) and brain natriuretic peptide (BNP)—both of which are commonly used markers for hypertrophy [65–67]. Importantly, there is a potential divergence between the signalling pathways involved in the cardioprotective response and the

hypertrophic response to urocortins. Namely, the p42/p44 MAPK pathway, though required for cardioprotection, does not appear to be involved in the hypertrophic effect of Ucn 1. Conversely, the PI3K/Akt pathway does appear to be required for it to induce hypertrophy [65]. In terms of practical application, it would be extremely helpful if a method is found to synthesize analogues of urocortins that are capable of activating selected protective pathways without any concurrent hypertrophic response.

A number of elegant studies have revealed a role for PKC ϵ in cardioprotection during cardiac ischemia/reperfusion injury (reviewed in [52]). The PKC family of kinases phosphorylates serine and threonine residues on target proteins. Eleven isozymes of PKC have been recognized so far and are classified on the basis of calcium dependency and regulation by lipid modulators [68]. However, the specificity of the PKC response to Ucn 1 exposure has been gleaned from microarray analyses that show elevated levels of PKCE in rat primary cardiomyocytes following 24 h exposure to Ucn 1, with no differences detected for other PKC isozymes; thus suggesting urocortins selectively activate PKCε [45,55,57,58]. When activated, PKCs translocate from the cytosol to the plasma membranes. This translocation occurs when cytosolic PKC binds to an isozymespecific membrane-bound anchor protein or RACK (receptor for activated C kinase) which in turn activates PKC isozyme transported to its target protein [58]. Small peptides of six to eight amino acids have been used to inhibit specific isozymes of PKC from binding to the specific RACK, as well as, pseudo-RACK peptides to enhance the function of specific PKCs. Pseudo RACKS specific for PKCε have shown that it acts as a cardioprotective agent [69-71]. Furthermore, evidence that PKCE is the downstream modulator of Ucn 1 was provided by the use of specific PKCE inhibitor peptides, that when introduced into cardiomyocytes, prior to simulated ischemia, resulted in the loss of Ucn 1 cardioprotective effect [58].

Interestingly, the PI3K/Akt and ERK1/2 kinase cardioprotective signalling pathways, in addition to PKC£ appear to be common to most cardioprotective peptide agents [50–52,72]. However, this does not exclude the possible involvement of other kinase pathways such as STAT-3, recently shown to be required for cardioprotection by the canonical Akt activator, insulin [73]. Furthermore, the involvement of other kinases known to be activated by urocortins, such as PKA, remains to be tested.

7. Other cardioprotective mechanisms of the CRH-peptide family

In addition to the activation of protein kinase signalling pathways, a significant contribution of Ucn 1-mediated cardioprotection involves *de novo* protein synthesis. Expression of the cardioprotective heat shock protein 90 (HSP90) has been reported to be induced by Ucn 1 and this effect is blocked by the MEK1/2 inhibitor PD98059 [74]. Moreover, the expression of cardiotrophin-1 (CT-1), an additional cardioprotective peptide, is enhanced upon exposure of cells to Ucn 1 [75].

Microarray profiling has helped dramatically to unravel the gene expression profile effects of Ucn 1 [55]. In this study the protein identified was the $K_{\rm ATP}$ channel. This channel opens in

response to a drop in the cytosolic concentration of ATP, and therefore it is regarded as a sensor of the metabolic state of a cell. These channels, when open during stressful stimuli such as ischemia/reperfusion are "cardioprotective" [55,57]. There are two known subtypes of K_{ATP} channel, Kir 6.1 and Kir 6.2, small transmembrane proteins that form the pore of the channel. For a KATP channel to be fully functional requires additional protein subunits - namely the sulphonylurea receptors (SUR). These SUR receptors are responsible for sensing and binding ATP/ADP and ultimately gating the channel pore [53,76]. Ucn 1 induces the expression of the Kir 6.1 potassium channel subunit in both primary cardiomyocytes and the whole heart [55]. Using an antagonist to the mitochondrial K_{ATP} channel, 5-hydroxydecanoate (5-HD) and dominant negative versions of Kir 6.1, the complete loss of the cardioprotective properties of Ucn 1 was demonstrated when Kir 6.1 was inhibited. Furthermore, openers of K_{ATP} channel, including cromakalim, are accordingly cardioprotective during simulated ischemia/reperfusion injury in vitro [55].

Ucn 1 also regulates iPLA₂ which belongs to a superfamily of phosphoplipases. iPLA₂ catalyses the breakdown of membrane phospholipids into arachidonic acid (AA), a precursor of prostaglandins, leukotrienes and a minor metabolite lysophosphatidylcholine (LPC). iPLA₂ catalytic activity is enhanced during ischemia/reperfusion, with a corresponding increase in its metabolites AA and LPC; the latter being highly cardiotoxic, and as such are implicated in cell death during ischemia/reperfusion injury [77,78]. Ucn 1, as well as the iPLA₂ inhibitor, bromoenol lactone (BEL), was found to lower the expression levels of LPC in cardiomyocytes and therefore protect cardiomyocytes against injury caused by LPC and ischemia/reperfusion to a similar degree [56,57].

In recent years the crucial role of the MPTP in reperfusion injury has become increasingly apparent. This inner mitochondrial channel opens in response to elevated mitochondrial calcium, oxidative stress and adenine nucleotide depletion—the exact conditions that occur early during reperfusion following a period of ischemia [79]. A recent study, using the Langendorff perfused heart model, has shown that Ucn 1 can decrease MPTP opening during reperfusion in the intact heart [45,54]. The role of the mitochondria was demonstrated using the 'hot dog' technique in which 2-[3H]deoxyglucose ("hot DOG") becomes entrapped within mitochondria that undergo mitochondria permeability transition (MPT). An acute, 30 min pre-treatment with Ucn 1 significantly reduced the loss of 2-[3H]deoxyglucose with an accompanying reduction in the degree of ischemia/reperfusion-induced MPTP and associated tissue damage in rat hearts [45]. How Ucn 1 blocks the opening of MPTP is still not fully understood although it seems that, via activation of PKCε, it decreases the burst of reactive oxygen species (ROS) upon reperfusion, and therefore greatly reduces one of the prime activators of MPTP. Both by using a redox-sensitive dye in isolated cells, and by quantifying protein carbonylation as an index of oxidatively damaged proteins in the intact heart, Ucn 1 was shown to decrease levels of oxidative stress during reperfusion, and this could be blocked by the PKC inhibitor chelerythrine [45]. However, further detailed studies are needed to explain the role of PKCε on the inhibition of MPTP and its precise mechanism of action.

Further research and a better understanding of the signalling mechanism of the urocortins in cardioprotection and their role in the reduction of cell death is of immense importance and will enhance our ability to focus on exogenous modulation of cardioprotection, a useful clinical strategy for cardiac damage.

8. Urocortin as a therapeutic agent

The therapeutic potential of the urocortin peptides has been recognized for some time now, both in regard to its powerful cardioprotective properties, and other favourable cardiovascular effects mentioned above. The pharmacokinetics have been investigated, and the plasma half-life of 50 μg Ucn 1 in humans was found to be 52 min [80]. One particularly interesting aspect of urocortin treatment of heart failure is the apparent lack of desensitization after prolonged treatment [41]. This would be worthy of further investigation in relation to cardioprotection, since most forms of cardioprotection currently being investigated – including the canonical form, ischemic preconditioning – are transient.

As mentioned previously, urocortins have various effects on the heart, some of which would be undesirable for a human therapeutic agent. Some divergent aspects of the signalling pathways by which urocortins lead to hypertrophy and/or cardioprotection have been identified [65,81], raising the possibility of dissecting out these separate actions. A further important consideration is the non-cardiac effects of CRH administration. For this reason Ucn 2 and Ucn 3 are generally regarded as having greater potential, since they have much less affinity for the CRH-R1 receptor responsible for most of the centrally mediated effects. In sheep, activation of the pituitary–adrenal axis was observed in response to Ucn 2 or Ucn 3, though only minimal effects were observed in rats, mice and humans, possibly because the peptides have less affinity to CRH-R1 in these species.

In addition to being effective in experimental systems, therapeutic agents must be beneficial in heart failure patients who are often already receiving multiple pharmaceuticals. Using their sheep model of heart failure, Rademaker et al. showed that co-treatment with Ucn 2 and an ACE inhibitor, captopril, in heart failure produced significantly greater improvements in hemodynamics, hormonal profile and renal function than achieved by captopril alone [82].

9. What does the future hold for the urocortins?

The wide range of biological effects mediated by the urocortins are far from being completely understood. For example, although it has been known for some years that mice deficient for CRH-R2 become hypervascularized postnatally [83], only recently has it been shown that inhibition of neo-vascularization by Ucn 2 overexpression can contribute to tumor growth inhibition in vivo [84]. Although an exciting finding in terms of cancer research, this suggests that the administration of Ucn 2 or its homologues may have detrimental effects in patients with heart failure or post-myocardial infarction, in which

neovascularization is desirable. Similarly, Ucn 2 has recently been shown to function as a local negative regulator of glucose uptake in skeletal muscle, suggesting that prolonged Ucn 2 treatment might alter blood glucose levels [16]. Long-term studies of urocortin administration in animal studies still seem warranted.

On the other hand, the strong preference that some members of the family exhibit for binding to CRH-R2, and the highly beneficial effects observed even after several days administration in sheep models of heart failure, does seem extremely promising. Of course, its effect in humans must be determined, and indeed, one company (Neurocrine Biosciences Inc.), has now completed a phase I clinical trial using Ucn 2, and plans to initiate phase II studies in patients with mild to moderate congestive heart disease in 2008. Clearly, the precise underlying molecular mechanisms by which they act still need to be fully described yet the in vitro, in vivo and ex vivo data presented demonstrate a bright future for the use of the urocortins as a therapeutic small molecule.

Acknowledgements

SMD is funded by the Medical Research Council, UK. PAT would like to thank the Biotechnology and Biological Sciences Research Council and the British Heart Foundation for research grant support and the Gerald Kerkut Charitable Trust for funding the PhD studentship of AER.

REFERENCE

- [1] Vaughan J, Donaldson C, Bittencourt J, Perrin MH, Lewis K, Sutton S, et al. Urocortin, a mammalian neuropeptide related to fish urotensin I and to corticotropin-releasing factor. Nature 1995;378:287–92.
- [2] Oki Y, Sasano H. Localization and physiological roles of urocortin. Peptides 2004;25:1745–9.
- [3] Kimura Y, Takahashi K, Totsune K, Muramatsu Y, Kaneko C, Darnel AD, et al. Expression of urocortin and corticotropin-releasing factor receptor subtypes in the human heart. J Clin Endocrinol Metab 2002;87:340–6.
- [4] Perrin M, Donaldson C, Chen R, Blount A, Berggren T, Bilezikjian L, et al. Identification of a second corticotropinreleasing factor receptor gene and characterization of a cDNA expressed in heart. Proc Natl Acad Sci USA 1995;92:2969–73.
- [5] Nishikimi T, Miyata A, Horio T, Yoshihara F, Nagaya N, Takishita S, et al. Urocortin, a member of the corticotropinreleasing factor family, in normal and diseased heart. Am J Physiol Heart Circ Physiol 2000;279:H3031–9.
- [6] Lovenberg TW, Liaw CW, Grigoriadis DE, Clevenger W, Chalmers DT, De Souza EB, et al. Cloning and characterization of a functionally distinct corticotropinreleasing factor receptor subtype from rat brain. Proc Natl Acad Sci USA 1995;92:836–40.
- [7] Hsu SY, Hsueh AJ. Human stresscopin and stresscopinrelated peptide are selective ligands for the type 2 corticotropin-releasing hormone receptor. Nat Med 2001;7:605–11.
- [8] Reyes TM, Lewis K, Perrin MH, Kunitake KS, Vaughan J, Arias CA, et al. Urocortin II: a member of the corticotropinreleasing factor (CRF) neuropeptide family that is

- selectively bound by type 2 CRF receptors. Proc Natl Acad Sci USA 2001;98:2843–8.
- [9] Lewis K, Li C, Perrin MH, Blount A, Kunitake K, Donaldson C, et al. Identification of urocortin III, an additional member of the corticotropin-releasing factor (CRF) family with high affinity for the CRF2 receptor. Proc Natl Acad Sci USA 2001;98:7570–5.
- [10] Li C, Chen P, Vaughan J, Lee KF, Vale W. Urocortin 3 regulates glucose-stimulated insulin secretion and energy homeostasis. Proc Natl Acad Sci USA 2007;104:4206–11.
- [11] Takahashi K, Totsune K, Murakami O, Saruta M, Nakabayashi M, Suzuki T, et al. Expression of urocortin III/ stresscopin in human heart and kidney. J Clin Endocrinol Metab 2004;89:1897–903.
- [12] Grace CR, Perrin MH, Cantle JP, Vale WW, Rivier JE, Riek R. Common and divergent structural features of a series of corticotropin releasing factor-related peptides. J Am Chem Soc 2007;129:16102–14.
- [13] Rivier J, Rivier C, Vale W. Synthetic competitive antagonists of corticotropin-releasing factor: effect on ACTH secretion in the rat. Science 1984;224:889–91.
- [14] Carlin KM, Vale WW, Bale TL. Vital functions of corticotropin-releasing factor (CRF) pathways in maintenance and regulation of energy homeostasis. Proc Natl Acad Sci USA 2006;103:3462–7.
- [15] Solinas G, Summermatter S, Mainieri D, Gubler M, Montani JP, Seydoux J, et al. Corticotropin-releasing hormone directly stimulates thermogenesis in skeletal muscle possibly through substrate cycling between de novo lipogenesis and lipid oxidation. Endocrinology 2006;147: 31–8.
- [16] Chen A, Brar B, Choi CS, Rousso D, Vaughan J, Kuperman Y, et al. Urocortin 2 modulates glucose utilization and insulin sensitivity in skeletal muscle. Proc Natl Acad Sci USA 2006;103:16580–5.
- [17] Rothwell NJ. CRF is involved in the pyrogenic and thermogenic effects of interleukin 1 beta in the rat. Am J Physiol 1989;256:E111–5.
- [18] Fekete EM, Zorrilla EP. Physiology, pharmacology, and therapeutic relevance of urocortins in mammals: ancient CRF paralogs. Front Neuroendocrinol 2007;28:1–27.
- [19] Spina M, Merlo-Pich E, Chan RK, Basso AM, Rivier J, Vale W, et al. Appetite-suppressing effects of urocortin, a CRFrelated neuropeptide. Science 1996;273:1561–4.
- [20] Cullen MJ, Ling N, Foster AC, Pelleymounter MA. Urocortin, corticotropin releasing factor-2 receptors and energy balance. Endocrinology 2001;142:992–9.
- [21] Kakiya S, Yokoi H, Arima H, Iwasaki Y, Oki Y, Oiso Y. Central administration of urocortin inhibits vasopressin release in conscious rats. Neurosci Lett 1998;248:144–6.
- [22] Lovejoy DA, Jahan S. Phylogeny of the corticotropinreleasing factor family of peptides in the metazoa. Gen Comp Endocrinol 2006;146:1–8.
- [23] Tsatsanis C, Androulidaki A, Dermitzaki E, Gravanis A, Margioris AN. Corticotropin releasing factor receptor 1 (CRF1) and CRF2 agonists exert an anti-inflammatory effect during the early phase of inflammation suppressing LPSinduced TNF-alpha release from macrophages via induction of COX-2 and PGE2. J Cell Physiol 2007;210:774–83.
- [24] Martinez V, Tache Y. CRF1 receptors as a therapeutic target for irritable bowel syndrome. Curr Pharm Des 2006;12:4071–88.
- [25] Wu Y, Zhou H, Xu Y, Li S. Enhanced expression of urocortin in lung tissues of rats with allergic asthma. Biochem Biophys Res Commun 2006;341:532–40.
- [26] Brar BK, Stephanou A, Okosi A, Lawrence KM, Knight RA, Marber MS, et al. CRH-like peptides protect cardiac myocytes from lethal ischaemic injury. Mol Cell Endocrinol 1999;158:55–63.

- [27] Knight RA, Chen-Scarabelli C, Yuan Z, McCauley RB, Di RJ, Scarabelli GM, et al. Cardiac release of urocortin precedes the occurrence of irreversible myocardial damage in the rat heart exposed to ischemia/reperfusion injury. FEBS Lett 2008;582:984–90.
- [28] Charles CJ, Rademaker MT, Richards AM, Yandle TG. Plasma urocortin 1 in sheep: regional sampling and effects of experimental heart failure. Peptides 2006;27:1801–5.
- [29] Ng LL, Loke IW, O'Brien RJ, Squire IB, Davies JE. Plasma urocortin in human systolic heart failure. Clin Sci (Lond) 2004;106:383–8.
- [30] Terui K, Higashiyama A, Horiba N, Furukawa KI, Motomura S, Suda T. Coronary vasodilation and positive inotropism by urocortin in the isolated rat heart. J Endocrinol 2001;169:177–83.
- [31] Coste SC, Kesterson RA, Heldwein KA, Stevens SL, Heard AD, Hollis JH, et al. Abnormal adaptations to stress and impaired cardiovascular function in mice lacking corticotropin-releasing hormone receptor-2. Nat Genet 2000:24:403-9.
- [32] Parkes DG, Vaughan J, Rivier J, Vale W, May CN. Cardiac inotropic actions of urocortin in conscious sheep. Am J Physiol 1997;272:H2115–22.
- [33] Charles CJ, Jardine DL, Nicholls MG, Rademaker MT, Richards AM. Urocortin 1 exhibits potent inhibition of cardiac sympathetic nerve activity in conscious sheep. J Hypertens 2008;26:53–60.
- [34] Rademaker MT, Charles CJ, Espiner EA, Frampton CM, Lainchbury JG, Richards AM. Endogenous urocortins reduce vascular tone and rennin–aldosterone/endothelin activity in experimental heart failure. Eur Heart J 2005;26:2046–54.
- [35] Rademaker MT, Charles CJ, Espiner EA, Fisher S, Frampton CM, Kirkpatrick CM, et al. Beneficial hemodynamic, endocrine, and renal effects of urocortin in experimental heart failure: comparison with normal sheep. J Am Coll Cardiol 2002;40:1495–505.
- [36] Rademaker MT, Cameron VA, Charles CJ, Richards AM. Integrated hemodynamic, hormonal, and renal actions of urocortin 2 in normal and paced sheep: beneficial effects in heart failure. Circulation 2005;112:3624–32.
- [37] Rademaker MT, Cameron VA, Charles CJ, Richards AM. Urocortin 3: haemodynamic, hormonal, and renal effects in experimental heart failure. Eur Heart J 2006;27:2088–98.
- [38] Bale TL, Hoshijima M, Gu Y, Dalton N, Anderson KR, Lee KF, et al. The cardiovascular physiologic actions of urocortin II: acute effects in murine heart failure. Proc Natl Acad Sci USA 2004;101:3697–702.
- [39] Yao X, He GW, Chan FL, Lau CW, Tsang SY, Chen ZY, et al. Endothelium-dependent and -independent coronary relaxation induced by urocortin. J Card Surg 2002;17: 347–9.
- [40] Grossini E, Molinari C, Mary DA, Marino P, Vacca G. The effect of urocortin II administration on the coronary circulation and cardiac function in the anaesthetized pig is nitric-oxide-dependent. Eur J Pharmacol 2008;578:242–8.
- [41] Rademaker MT, Charles CJ, Espiner EA, Frampton CM, Lainchbury JG, Richards AM. Four-day urocortin-I administration has sustained beneficial haemodynamic, hormonal, and renal effects in experimental heart failure. Eur Heart J 2005;26:2055–62.
- [42] Rademaker MT, Charles CJ, Richards AM. Urocortin 1 administration from onset of rapid left ventricular pacing represses progression to overt heart failure. Am J Physiol Heart Circ Physiol 2007;293:H1536–44.
- [43] Davis ME, Pemberton CJ, Yandle TG, Fisher SF, Lainchbury JG, Frampton CM, et al. Urocortin 2 infusion in human heart failure. Eur Heart J 2007;28:2589–97.
- [44] Davis ME, Pemberton CJ, Yandle TG, Fisher SF, Lainchbury JG, Frampton CM, et al. Urocortin 2 infusion in healthy

- humans: hemodynamic, neurohormonal, and renal responses. J Am Coll Cardiol 2007;49:461–71.
- [45] Townsend PA, Davidson SM, Clarke SJ, Khaliulin I, Carroll CJ, Scarabelli TM, et al. Urocortin prevents mitochondrial permeability transition in response to reperfusion injury indirectly by reducing oxidative stress. Am J Physiol Heart Circ Physiol 2007;293:H928–38.
- [46] Okosi A, Brar BK, Chan M, D'Souza L, Smith E, Stephanou A, et al. Expression and protective effects of urocortin in cardiac myocytes. Neuropeptides 1998;32:167–71.
- [47] Brar BK, Jonassen AK, Egorina EM, Chen A, Negro A, Perrin MH, et al. Urocortin-II and urocortin-III are cardioprotective against ischemia reperfusion injury: an essential endogenous cardioprotective role for corticotropin releasing factor receptor type 2 in the murine heart. Endocrinology 2004;145:24–35.
- [48] Brar BK, Jonassen AK, Stephanou A, Santilli G, Railson J, Knight RA, et al. Urocortin protects against ischemic and reperfusion injury via a MAPK-dependent pathway. J Biol Chem 2000;275:8508–14.
- [49] Scarabelli TM, Pasini E, Stephanou A, Comini L, Curello S, Raddino R, et al. Urocortin promotes hemodynamic and bioenergetic recovery and improves cell survival in the isolated rat heart exposed to ischemia/reperfusion. J Am Coll Cardiol 2002;40:155–61.
- [50] Schulz R, Cohen MV, Behrends M, Downey JM, Heusch G. Signal transduction of ischemic preconditioning. Cardiovasc Res 2001;52:181–98.
- [51] Hausenloy DJ, Yellon DM. Survival kinases in ischemic preconditioning and postconditioning. Cardiovasc Res 2006;70:240–53.
- [52] Downey JM, Davis AM, Cohen MV. Signaling pathways in ischemic preconditioning. Heart Fail Rev 2007;12:181–8.
- [53] Lawrence KM, Latchman DS. The Urocortins: mechanisms of cardioprotection and therapeutic potential. Mini Rev Med Chem 2006;6:1119–26.
- [54] Baines CP. The mitochondrial permeability transition pore as a target of cardioprotective signalling. Am J Physiol Heart Circ Physiol 2007;293:H903–4.
- [55] Lawrence KM, Chanalaris A, Scarabelli T, Hubank M, Pasini E, Townsend PA, et al. K(ATP) channel gene expression is induced by urocortin and mediates its cardioprotective effect. Circulation 2002;106:1556–62.
- [56] Lawrence KM, Scarabelli TM, Turtle L, Chanalaris A, Townsend PA, Carroll CJ, et al. Urocortin protects cardiac myocytes from ischemia/reperfusion injury by attenuating calcium-insensitive phospholipase A2 gene expression. FASEB J 2003;17:2313–5.
- [57] Lawrence KM, Townsend PA, Davidson SM, Carroll CJ, Eaton S, Hubank M, et al. The cardioprotective effect of urocortin during ischaemia/reperfusion involves the prevention of mitochondrial damage. Biochem Biophys Res Commun 2004;321:479–86.
- [58] Lawrence KM, Kabir AM, Bellahcene M, Davidson S, Cao XB, McCormick J, et al. Cardioprotection mediated by urocortin is dependent on PKCepsilon activation. FASEB J 2005;19:831–3.
- [59] Schulman D, Latchman DS, Yellon DM. Urocortin protects the heart from reperfusion injury via upregulation of p42/p44 MAPK signaling pathway. Am J Physiol Heart Circ Physiol 2002;283:H1481–8.
- [60] Latchman DS. Urocortin protects against ischemic injury via a MAPK-dependent pathway. Trends Cardiovasc Med 2001;11:167–9.
- [61] Brar BK, Stephanou A, Knight R, Latchman DS. Activation of protein kinase B/Akt by urocortin is essential for its ability to protect cardiac cells against hypoxia/ reoxygenation-induced cell death. J Mol Cell Cardiol 2002;34:483–92.

- [62] Zha J, Harada H, Yang E, Jockel J, Korsmeyer SJ. Serine phosphorylation of death agonist BAD in response to survival factor results in binding to 14-3-3 not BCL-X(L). Cell 1996;87:619–28.
- [63] Cardone MH, Roy N, Stennicke HR, Salvesen GS, Franke TF, Stanbridge E, et al. Regulation of cell death protease caspase-9 by phosphorylation. Science 1998;282:1318–21.
- [64] Terada K, Kaziro Y, Satoh T. Analysis of Ras-dependent signals that prevent caspase-3 activation and apoptosis induced by cytokine deprivation in hematopoietic cells. Biochem Biophys Res Commun 2000;267:449–55.
- [65] Chanalaris A, Lawrence KM, Townsend PA, Davidson S, Jamshidi Y, Stephanou A, et al. Hypertrophic effects of urocortin homologous peptides are mediated via activation of the Akt pathway. Biochem Biophys Res Commun 2005;328:442–8.
- [66] Railson JE, Liao Z, Brar BK, Buddle JC, Pennica D, Stephanou A, et al. Cardiotrophin-1 and urocortin cause protection by the same pathway and hypertrophy via distinct pathways in cardiac myocytes. Cytokine 2002;17:243–53.
- [67] Barry SP, Davidson SM, Townsend PA. Molecular regulation of cardiac hypertrophy. Int J Biochem Cell Biol 2008;40:2023–39.
- [68] Naruse K, King GL. Protein kinase C and myocardial biology and function. Circ Res 2000;86:1104–6.
- [69] Mackay K, Mochly-Rosen D. Localization, anchoring, and functions of protein kinase C isozymes in the heart. J Mol Cell Cardiol 2001;33:1301–7.
- [70] Souroujon MC, Mochly-Rosen D. Peptide modulators of protein-protein interactions in intracellular signaling. Nat Biotechnol 1998;16:919–24.
- [71] Chen L, Wright LR, Chen CH, Oliver SF, Wender PA, Mochly-Rosen D. Molecular transporters for peptides: delivery of a cardioprotective epsilonPKC agonist peptide into cells and intact ischemic heart using a transport system, R(7). Chem Biol 2001;8:1123–9.
- [72] Hausenloy DJ, Yellon DM. New directions for protecting the heart against ischaemia-reperfusion injury: targeting the reperfusion injury salvage kinase (RISK)-pathway. Cardiovasc Res 2004;61:448–60.
- [73] Fuglesteg BN, Suleman N, Tiron C, Kanhema T, Lacerda L, Andreasen TV, et al. Signal transducer and activator of transcription 3 is involved in the cardioprotective signalling pathway activated by insulin therapy at reperfusion. Basic Res Cardiol 2008;103:444–53.
- [74] Brar BK, Railson J, Stephanou A, Knight RA, Latchman DS. Urocortin increases the expression of heat shock protein 90 in rat cardiac myocytes in a MEK1/2-dependent manner. J Endocrinol 2002;172:283–93.
- [75] Janjua S, Lawrence KM, Ng LL, Latchman DS. The cardioprotective agent urocortin induces expression of CT-1. Cardiovasc Toxicol 2003;3:255–62.
- [76] Inagaki N, Tsuura Y, Namba N, Masuda K, Gonoi T, Horie M, et al. Cloning and functional characterization of a novel

- ATP-sensitive potassium channel ubiquitously expressed in rat tissues, including pancreatic islets, pituitary, skeletal muscle, and heart. J Biol Chem 1995;270:5691–4.
- [77] Cummings BS, McHowat J, Schnellmann RG. Phospholipase A(2)s in cell injury and death. J Pharmacol Exp Ther 2000;294:793–9.
- [78] Daleau P. Lysophosphatidylcholine, a metabolite which accumulates early in myocardium during ischemia, reduces gap junctional coupling in cardiac cells. J Mol Cell Cardiol 1999;31:1391–401.
- [79] Halestrap AP, Clarke SJ, Javadov SA. Mitochondrial permeability transition pore opening during myocardial reperfusion—a target for cardioprotection. Cardiovasc Res 2004;61:372–85.
- [80] Davis ME, Pemberton CJ, Yandle TG, Lainchbury JG, Rademaker MT, Nicholls MG, et al. Urocortin-1 infusion in normal humans. J Clin Endocrinol Metab 2004;89:1402–9.
- [81] Davidson SM, Townsend PA, Carroll C, Yurek-George A, Balasubramanyam K, Kundu TK, et al. The transcriptional coactivator p300 plays a critical role in the hypertrophic and protective pathways induced by phenylephrine in cardiac cells but is specific to the hypertrophic effect of urocortin. Chembiochem 2005;6:162–70.
- [82] Rademaker MT, Charles CJ, Nicholls MG, Richards AM. Urocortin 2 combined with angiotensin-converting enzyme inhibition in experimental heart failure. Clin Sci (Lond) 2008;114:635–42.
- [83] Bale TL, Giordano FJ, Hickey RP, Huang Y, Nath AK, Peterson KL, et al. Corticotropin-releasing factor receptor 2 is a tonic suppressor of vascularization. Proc Natl Acad Sci USA 2002;99:7734–9.
- [84] Hao Z, Huang Y, Cleman J, Jovin IS, Vale WW, Bale TL, et al. Urocortin2 inhibits tumor growth via effects on vascularization and cell proliferation. Proc Natl Acad Sci USA 2008;105:3939–44.
- [85] Parkes DG, May CN. Urocortin: a novel player in cardiac control. News Physiol Sci 2000;15:264–8.
- [86] Wu Y, Xu Y, Zhou H, Tao J, Li S. Expression of urocortin in rat lung and its effect on pulmonary vascular permeability. J Endocrinol 2006;189:167–78.
- [87] Uzuki M, Sasano H, Muramatsu Y, Totsune K, Takahashi K, Oki Y, et al. Urocortin in the synovial tissue of patients with rheumatoid arthritis. Clin Sci (Lond) 2001;100:577–89.
- [88] Muramatsu Y, Sugino N, Suzuki T, Totsune K, Takahashi K, Tashiro A, et al. Urocortin and corticotropin-releasing factor receptor expression in normal cycling human ovaries. J Clin Endocrinol Metab 2001;86:1362–9.
- [89] Li X, Hu J, Zhang R, Sun X, Zhang Q, Guan X, et al. Urocortin ameliorates diabetic nephropathy in obese db/db mice. Br J Pharmacol 2008;154:1025–34.
- [90] Wang J, Xu Y, Xu Y, Zhu H, Zhang R, Zhang G, et al. Urocortin's inhibition of tumor growth and angiogenesis in hepatocellular carcinoma via corticotrophin-releasing factor receptor 2. Cancer Invest 2008;26:359–68.